

NOTES

TRISARUBICINOL, NEW ANTITUMOR ANTHRACYCLINE ANTIBIOTIC

AKIHIRO YOSHIMOTO, YASUE MATSUZAWA, YOSHIYUKI MATSUSHITA and TOSHIKAZU OKI

Central Research Laboratories, Sanraku-Ocean Co., Ltd., 4-9-1 Johnan, Fujisawa 251, Japan

TOMIO TAKEUCHI and HAMA O UMEZAWA

Institute of Microbial Chemistry, 3-14-23 Kamiosaki, Shinagawa-ku, Tokyo 141, Japan

(Received for publication July 17, 1981)

As previously reported¹⁻⁴⁾, an aclacinomycin-negative mutant strain KE303, which was isolated from *Streptomyces galilaeus* MA144-M1, is capable of producing new potent antitumor anthracycline antibiotics by the microbial conversion of biologically inactive anthracyclines. In this communication, we describe the microbial glycosidation of carminomycinone and 13-dihydrocarminomycinone using strain KE303, and the characterization of 13-dihydrocarminomycinone trisaccharide, trisarubicinol (Fig. 1).

The strain KE303 was cultured in 500-ml flasks containing 50 ml medium of the following composition: Soluble starch 1.5 %, glucose 1 %, soy bean meal (Ajinomoto Co.) 2 %, yeast extract 0.2 %, NaCl 0.3 %, K_2HPO_4 0.1 %, $MgSO_4 \cdot 7H_2O$ 0.1 %, $CuSO_4 \cdot 5H_2O$ 0.0007 %, $FeSO_4 \cdot 7H_2O$ 0.0001 %, $MnCl_2 \cdot 4H_2O$ 0.0008 %, $ZnSO_4 \cdot 7H_2O$ 0.0002 %, pH 7.4. The cultivation was carried out for 17 hours at 28°C on a rotary shaker, and then 0.5 ml of methanol solution of carminomycinone (1 mg/ml) was added to each flask at the concentration of 10 µg/ml, and the cultivation was further continued for 24 hours to complete the glycosidation. The pigments were extracted from the cultured broth (5 ml) with a solvent mixture of chloroform - methanol (3:2, v/v, 5 ml), and the chloroform layer was concentrated to dryness, dissolved in 0.2 ml of chloroform, spotted 20 µl onto silica gel thin-layer (F_{254} plate, E. Merck Co.) and developed with chloroform - methanol - aqueous ammonia (100:10:0.3, v/v/v) mixture. After drying, the spots corresponding to trisarubicinol (I) (Rf=

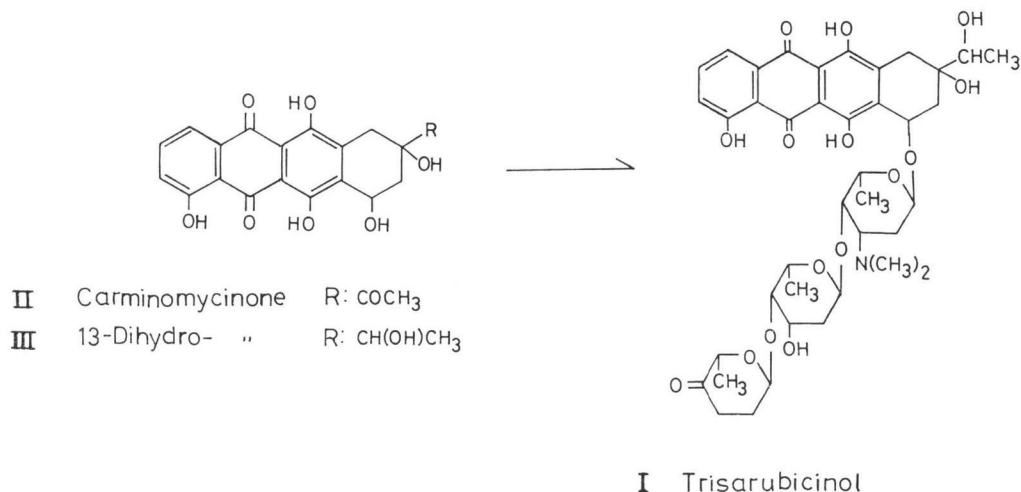
0.79), residual carminomycinone (II) (Rf=0.67) and its reduced form, 13-dihydrocarminomycinone (III) (Rf=0.43) were determined by a Shimadzu chromatoscanner model CS-910. The conversion rate of II to I was about 30 %.

Fifty liters of the cultured broth were centrifuged to harvest the mycelium, and the pigments were extracted from the mycelium with 8 liters of acetone, concentrated to one-third volume, and re-extracted with 3 liters of chloroform. After concentration to dryness, the crude pigmented residue was dissolved in 100 ml of chloroform - methanol (1:2, v/v), subjected to Sephadex LH-20 column, and eluted with the same solvent mixture. The initial red eluate was concentrated, dissolved in a small amount of chloroform, and chromatographed on preparative thin-layer (Kieselgel 60PF₂₅₄, E. Merck Co.) using a chloroform - methanol (20:1, v/v) mixture. The major band corresponding to I showing Rf at 0.2 was scraped off, and extracted with 200 ml of chloroform - methanol - aqueous ammonia (100:15:0.2, v/v/v) mixture. The extract was concentrated to dryness, dissolved in 0.1 M acetate buffer (pH 3.5) and washed with toluene. The aqueous layer was neutralized the pH to 7.0 with sodium bicarbonate and extracted with chloroform. The chloroform extract was dried over sodium sulfate and concentrated to 2.5 ml under reduced pressure. To the concentrate excess *n*-hexane was added to form a dark red precipitate, and 42 mg of pure trisarubicinol were obtained by filtration and drying of the precipitate *in vacuo*.

Physicochemical properties of I are as follows: m.p. 149~152°C; IR (KBr) cm^{-1} : 1720, 1600, 1290, 1005; $\lambda_{max}^{90\%MeOH}$ nm ($E_{1cm}^{1\%}$): 234(419), 254(342), 292(95), 465(125), 493(162), 514(20), 526(117), 575(15); $\lambda_{max}^{0.1NNaOH-90\%MeOH}$ nm ($E_{1cm}^{1\%}$): 241(482), 286(92), 560(181), 596(154); $[\alpha]_D^{25} + 1.84^\circ$ (*c* 0.8, $CHCl_3$); Anal. Calcd. for $C_{40}H_{51}NO_{15}$ (m.w. 785.84); C 61.14, H 6.54, N 1.78, O 30.54; Found: C 60.62, H 6.47, N 1.93%.

The PMR spectrum of I exhibited four signals in the anomeric region at δ 5.03, 5.07, 5.24 and 5.52 which were assigned to the C-1'', C-1''', C-7 and C-1' protons, respectively. This indicated that I had trisaccharide moiety. The signal at δ 2.39 showing the acetyl protons at C-9 of car-

Fig. 1. Microbial glycosidation of carminomycinone and 13-dihydrocarminomycinone by a mutant strain KE303 of *Streptomyces galilaeus* MA144-M1.



minomycinone, which was used as the substrate, was missing in the PMR spectrum of I.

The structure of aglycone moiety obtained by acid hydrolysis of I in 0.1 N HCl at 85°C for 30 minutes was identified as 13-dihydrocarminomycinone in direct comparison with the melting point, R_f value, IR and mass spectra of the authentic sample⁵⁾. On the other hand, sugar moieties obtained from the above hydrolysate were identified to be composed of rhodosamine, 2-deoxyfucose and cinerulose A by silica gel TLC as detected in the acid hydrolysate of aclacinomycin A⁶⁾. The sequence of sugar moiety was examined to be cinerulosyl-2-deoxyfucosyl-rhodosaminide by partial methanolysis in 0.01 N methanolic hydrogen chloride-acetone mixture at room temperature for 45 minutes, as previously described. The possible site for glycosidic linkage of 13-dihydrocarminomycinone should be the hydroxyl group at C-7 position. In the CMR spectrum of I, the signal at C-7 (δ 70.0) indicated that the glycosidic moiety linked to the C-7 position of 13-dihydrocarminomycinone.

The antibiotic I showed a marked antitumor activity against murine leukemia L1210 in CDF₁ mice. When 7.5 and 5.0 mg/kg/day of I was administered intraperitoneally once daily for 10 days, the increase of life span was 143 and 114 %, respectively. The concentrations required to inhibit synthesis by 50 % (IC₅₀) of DNA and RNA indicated that trisarubicinol was a more potent inhibitor of RNA synthesis than was car-

Table 1. Inhibition of the cell growth and macromolecular synthesis of cultured L1210 leukemia cells by trisarubicinol and carminomycin I.

Anthracycline	IC ₅₀ (μ M)			Ratio
	Growth	DNA synthesis	RNA synthesis	$\frac{IC_{50} \text{ DNA}}{IC_{50} \text{ RNA}}$
Trisarubicinol	0.013	0.48	0.08	6.0
Carminomycin I	0.020	0.39	0.56	0.7

IC₅₀ values were estimated by Probit analysis.

Cytotoxicity was determined on the day 2 culture.

minomycin I, as shown in Table 1. The IC₅₀ value of I for RNA synthesis was one-sixth that of DNA synthesis, while the IC₅₀ of carminomycin I for RNA and DNA syntheses was about equal.

References

- 1) YOSHIMOTO, A.; Y. MATSUZAWA, T. OKI, H. NAGANAWA, T. TAKEUCHI & H. UMEZAWA: Microbial conversion of ϵ -pyrromycinone and ϵ -isorhodomycinone to 1-hydroxy-13-dihydrodaunomycin and N-formyl-1-hydroxy-13-dihydrodaunomycin and their bioactivities. *J. Antibiotics* 33: 1150~1157, 1980
- 2) OKI, T.; A. YOSHIMOTO, Y. MATSUZAWA, T. TAKEUCHI & H. UMEZAWA: Biosynthesis of anthracycline antibiotics by *Streptomyces galilaeus*. I. Glycosidation of various anthra-

- cyclinones by an aclacinomycin-negative mutant and biosynthesis of aclacinomycins from akla-
vinone. J. Antibiotics 33: 1331~1340, 1980
- 3) YOSHIMOTO, A.; T. OKI, T. TAKEUCHI & H. UMEZAWA: Microbial conversion of anthra-
cyclinones to daunomycin by blocked mutants
of *Streptomyces coeruleorubidus*. J. Antibio-
tics 33: 1158~1166, 1980
 - 4) OKI, T.; A. YOSHIMOTO, Y. MATSUZAWA, T.
TAKEUCHI & H. UMEZAWA: New anthracy-
cline antibiotic, 2-hydroxyaclacinomycin A. J.
Antibiotics 34: 916~918, 1981
 - 5) MATSUZAWA, Y.; A. YOSHIMOTO, K. KOUNO &
T. OKI: Baumycin analogs isolated from
Actinomadura sp. J. Antibiotics 34: 774~776,
1981
 - 6) OKI, T.; I. KITAMURA, Y. MATSUZAWA, N.
SHIBAMOTO, T. OGASAWARA, A. YOSHIMOTO, T.
INUI, H. NAGANAWA, T. TAKEUCHI & H. UME-
ZAWA: Antitumor anthracycline antibiotics,
aclacinomycin A and analogues. II. Struc-
tural determination. J. Antibiotics 32: 801~
819, 1979